Tetrahedron Letters 51 (2010) 3146-3148

Contents lists available at ScienceDirect

Tetrahedron Letters

journal homepage: www.elsevier.com/locate/tetlet

## A mild and efficient synthetic protocol for Ferrier azaglycosylation promoted by ZnCl<sub>2</sub>/Al<sub>2</sub>O<sub>3</sub>

Feiging Ding, Ronny William, Bala Kishan Gorityala, Jimei Ma, Siming Wang, Xue-Wei Liu\*

Division of Chemistry and Biological Chemistry, School of Physical and Mathematical Sciences, Nanyang Technological University, Singapore 637371, Singapore

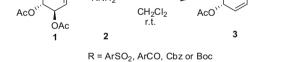
ARTICLE INFO	A B S T R A C T
Article history:	An improved method for the Ferrier sulfonamidoglycosylation of tri-O-acetyl-D-glucal with different <i>N</i> -
Received 11 February 2010	nucleophiles has been developed. $ZnCl_2$ impregnated on activated alumina acts as an excellent reagent
Revised 20 March 2010	system for the conversion of 3,4,6-tri-O-acetyl-D-glucal into 2,3-unsaturated- <i>N</i> -pseudoglycals with good
Accepted 12 April 2010	yields and preferential $\alpha$ -anomeric selectivity.
Available online 18 April 2010	© 2010 Elsevier Ltd. All rights reserved.

Ferrier rearrangement is a prominent reaction for the direct conversion of 3.4.6-tri-O-acetyl-p-glucal into 2.3-unsaturated glvcosides.<sup>1</sup> Development of this method is of broad interest due to the considerable importance of 2,3-unsaturated glycosides, which serve as basic building blocks. In general, Ferrier glycosylation reactions occur when glucals are treated with nucleophiles such as carbon nucleophiles, oxygen and/or sulfur nucleophiles (alcohols and thiols) in the presence of Lewis acids or oxidants.<sup>2</sup> On the other hand, nitrogen nucleophiles have not been used extensively in the Ferrier rearrangement.<sup>3</sup> Azide is the typical nucleophile used to afford the N-pseudoglycals.<sup>4</sup> This prompted us to develop a mild protocol for the conversion of 3,4,6-tri-O-acetyl-Dglucal into N-pseudoglycals with sulfonamides, carbamates, amides and azides. Nucleosides having an N-glycosidic linkage represent highly valuable pharmacological agents such as antibiotic, antineoplastic and antiviral compounds.<sup>5</sup> Subsequent modification of the unsaturated part of the pseudoglycals via epoxidation, hydroxylation, hydrogenation or aminohydroxylation further enhances the synthetic utility of this conversion.

To date, there are only two known examples of the Ferrier sulfonamidoglycosylation of glycals in the presence of  $B(C_6F_5)_3^6$  and  $BF_3 \cdot Et_2O^7$  as catalysts, respectively. Our group has been interested in new glycosylation techniques.<sup>8</sup> Recently, we demonstrated that the ZnCl<sub>2</sub>/Al<sub>2</sub>O<sub>3</sub> catalyzed Ferrier rearrangement of glucals with alcohols proceeds with high anomeric selectivity and in short reaction times.<sup>9</sup> To the best of our knowledge, the use of ZnCl<sub>2</sub>/Al<sub>2</sub>O<sub>3</sub> to promote the Ferrier sulfonamidoglycosylation of glycals has not been reported. Herein, we describe the synthesis of N-pseudoglycals by reaction of tri-O-acetyl-p-glucal (1) with nitrogen nucleophiles catalyzed by ZnCl<sub>2</sub>/Al<sub>2</sub>O<sub>3</sub> (Scheme 1).

Initially, a comparative study was carried out using tri-O-acetyl-D-glucal (1) and TsNH<sub>2</sub> as a model system with different Lewis acids (Table 1). Amongst the different catalysts tested, ZnCl<sub>2</sub>/

Al<sub>2</sub>O<sub>3</sub><sup>10</sup> (Table 1, entry 10) was found to be superior in terms of reaction time, vield, reaction profile and selectivity. Moreover, the use of Cu(OTf)<sub>2</sub>, Yb(OTf)<sub>2</sub>, IrCl<sub>3</sub>, RuCl<sub>3</sub> and ZnCl<sub>2</sub> as Lewis acid catalysts afforded the desired pseudoglycal products, albeit in longer reaction times, and moderate conversions and selectivities (Table 1, entries 1, 5 and 7-9). On the other hand, treatment of



ZnCl<sub>2</sub>/Al<sub>2</sub>O

Scheme 1. A synthetic model for the Ferrier azaglycosylation.

Table 1 Optimization of the catalyst for the Ferrier rearrangement<sup>a</sup>

RNH

Entry	Catalyst	Time (min)	Conversion (%) <sup>b</sup>	<b>α:</b> β <sup>c</sup>
1	Cu(OTf) <sub>2</sub>	120	100	77:23
2	FeCl <sub>3</sub>	120	Complex mixture	_
3	FeCl <sub>2</sub>	120	Complex mixture	_
4	Sn(OTf) <sub>2</sub>	180	Trace	_
5	Yb(OTf) <sub>2</sub>	120	100	68:32
6	PtCl <sub>2</sub>	180	No reaction	-
7	IrCl <sub>3</sub>	120	91	73:27
8	RuCl <sub>3</sub>	120	100	65:35
9	ZnCl <sub>2</sub>	120	92	62:38
10	ZnCl <sub>2</sub> /Al <sub>2</sub> O <sub>3</sub> <sup>d</sup>	20	100	88:12

<sup>a</sup> Reaction conditions: tri-O-acetyl-D-glucal (1) (100 mg, 0.37 mmol), TsNH<sub>2</sub> (1 equiv), catalyst (10 mol %), CH2Cl2 (1 mL).

<sup>b</sup> Determined by analysis of the <sup>1</sup>H NMR spectrum of the reaction mixture.

<sup>c</sup> The anomeric ratio was determined by integration of the anomeric hydrogen in the  $^1H$  NMR spectrum.  $^{\ \ d}$  ZnCl\_2 (1 equiv) on Al\_2O\_3 (5.3 equiv) was used per 1 equiv of tri-O-acetyl-D-

glucal.





<sup>\*</sup> Corresponding author. Tel.:+65 6316 8901; fax: +65 6791 1961. E-mail address: xuewei@ntu.edu.sg (X.-W. Liu).

<sup>0040-4039/\$ -</sup> see front matter © 2010 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetlet.2010.04.045

<i>F</i> .	Ding et	al./	/Tetrahedron	Letters 51	(2010)	3146-3148
------------	---------	------	--------------	------------	--------	-----------

Table 2	
---------	--

Scope of the Ferrier azaglycosylation of tri-O-acetyl-D-glucals	p-glucals <sup>11</sup>
---	-------------------------

Entry	Nucleophile	Product	Yield <sup>a</sup> (%)	$\alpha$ : $\beta^{b}$
1	2a O S, NH <sub>2</sub> O 2a	Aco Aco 3a	96	88:12
2	O <sub>2</sub> N 2b	AcO AcO AcO 3b	84	90:10
3	0, NH <sub>2</sub> , Zc	$\begin{array}{c} A_{CO} \\ A_{CO} \\ \end{array} \\ \begin{array}{c} O \\ A_{CO} \\ \end{array} \\ \begin{array}{c} O \\ 3c \end{array} \\ \begin{array}{c} H \\ N \\ S \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ O \\ O \\ O \\ O \\ \end{array} \\ \begin{array}{c} O \\ \end{array} \\ \begin{array}{c} O \\ O $	80	79:21 <sup>11</sup>
4	CI Zd		83	85:15 <sup>12</sup>
5	F 2e	AcO AcO AcO Be F	88	91:9 <sup>13</sup>
6	MeO 2f	ACO O H O ACO S O O ACO 3f OMe	91	80:20 <sup>14</sup>
7	°,'S,'NH₂ ⊙ 2g		90	91:9
8	0, NH₂ ∽S, NH₂ O 2h		88	85:15
9	NH <sub>2</sub>		54	74:26 <sup>15</sup>
10	2j NH <sub>2</sub>	n.r. <sup>c</sup>	-	-
11	CbzNH <sub>2</sub> <b>2k</b>	$A_{cO}$ $N_{cDz}$ $A_{cO}$ $3k$ $H_{cDz}$	80	92:8
12	BocNH <sub>2</sub> <b>21</b>	AcO AcO'	70	84:16
13	TMSN <sub>3</sub> 2m	AcO AcO 3m	92	86:14

<sup>a</sup> Isolated yields of anomeric mixtures after purification.
 <sup>b</sup> The anomeric ratio was determined by integration of the anomeric hydrogen in the <sup>1</sup>H NMR spectrum.

<sup>c</sup> No reaction.

glucal 1 with  $TsNH_2$  in the presence of 10 mol % of  $FeCl_3$  or  $FeCl_2$  led to a complex mixture of products (Table 1, entries 2 and 3),

while with  $Sn(OTf)_2$  and  $PtCl_2$  no reaction was observed (Table 1, entries 4 and 6), even after prolonged reaction times.

Using the optimized reaction conditions, the scope of the Ferrier azaglycosylation promoted by ZnCl<sub>2</sub>/Al<sub>2</sub>O<sub>3</sub> was examined. The results are summarized in Table 2. Generally, the reactions provided 2,3-unsaturated pseudoglycals in high to excellent yields with good anomeric selectivities. For example, sulfonamides 2a and 2f bearing electron-donating groups gave the corresponding pseudoglycals 3a and 3f in excellent yields and selectivities (Table 2, entries 1 and 6), while sulfonamides 2b-e bearing electronwithdrawing groups provided lower yields of the desired products, but typically with slightly higher selectivities (Table 2, entries 2-5). In addition, the aromatic sulfonamide [benzenesulfonamide (2g)] and methanesulfonamide (2h) (MsNH<sub>2</sub>) proved to be excellent nucleophiles providing the corresponding products in high yields and selectivities (Table 2, entries 7 and 8). Interestingly, extension of this reaction to other nitrogen nucleophiles such as benzamide **2i** furnished the corresponding pseudoglycal **3i**, in moderate vield and selectivity (Table 2, entry 9). To our knowledge. this is the first attempt using benzamide as a nucleophile in Ferrier glycosylation with tri-O-acetyl-D-glucal (1). This result encouraged us to further exploit the Ferrier azaglycosylation with benzyl carbamate (CbzNH<sub>2</sub>), *t*-butyl carbamate (BocNH<sub>2</sub>) and trimethylsilyl azide (TMSN<sub>3</sub>) as nucleophiles. Reaction of carbamates 2k and 2l with glucal 1 furnished the corresponding pseudoglycals 3k and **31**, in good yields and selectivities (Table 2, entries 11 and 12). These products can be easily transformed into the corresponding amines by removal of the protecting group. Likewise, the glycosylation of glucal 1 with TMSN<sub>3</sub> gave the corresponding glycosyl azide in 92% yield (Table 2, entry 13). In contrast, reaction of benzylamine 2j with tri-O-acetyl-D-glucal (1) was unsuccessful (Table 2, entry 10).

In summary, we have demonstrated a new protocol for the Ferrier azaglycosylation with various nitrogen nucleophiles under the influence of ZnCl<sub>2</sub>/Al<sub>2</sub>O<sub>3</sub>. The main advantages of this method are high anomeric selectivity and short reaction times. Low cost reagents and no aqueous work-up are required, and the catalyst could be recycled (up to three times).

## Acknowledgements

Financial support from NTU (RG50/08) and the Ministry of Education (MOE 2009-T2-1-030) Singapore is gratefully acknowledged.

## **References and notes**

- (a) Ferrier, R. J.; Prasad, N. J. Chem. Soc. **1969**, C, 570; (b) Ferrier, R. J.; Zubkov, O. A. Org. React. **2003**, 62, 569. and references therein; (c) Ferrier, R. J. Top. Curr. Chem. **2001**, 215, 153.
- 2. (a) Descotes, G.; Martin, J. C. Carbohydr. Res. 1977, 56, 168; (b) Klaffke, W.; Pudlo, P.; Springer, D.; Thiem, J. L. Ann. Chem. 1991, 6, 509; (c) Bhate, P.; Harton, D.; Priebe, W. Carbohydr. Res. 1985, 144, 331; (d) Babu, B. S.; Balsubramanian, K. K. Tetrahedron Lett. 2000, 41, 1271; (e) Das, S. K.; Reddy, K. A.; Roy, J. Synlett 2003, 1607; (f) Takhi, M.; Rahman, A.; Schmidt, R. R. Tetrahedron Lett. 2001, 42, 4053; (g) Masson, C.; Soto, J.; Besodes, M. Synlett 2000, 1281; (h) Yadav, J. S.; Reddy, B. V. S.; Chandraiah, L.; Reddy, K. S. Carbohydr. Res. 2001, 332, 221; (i) Swamy, N. R.; Venkateswarlu, A. Synthesis 2002, 598; (j) Bettadaiah, B. K.; Srinivas, P. Tetrahedron Lett. 2003, 44, 7257; (k) Yadav, J. S.; Reddy, B. V.; Reddy, J. S. J. Chem. Soc., Perkin Trans. 1 2002, 2390; (1) Yadav, J. S.; Reddy, B. V.; Murthy, C. V.; Kumar, G. M. Synlett 2000, 1450; (m) Smitha, G.; Reddy, S. C. Synthesis 2004, 834; (n) Gorityala, B. K.; Cai, S.; Lorpitthaya, R.; Ma, J.; Pasunooti, K. K.; Liu, X. W. Tetrahedron Lett. **2009**, *50*, 676; (o) Zhang, G.; Liu, Q.; Shi, L.; Wang, J. Tetrahedron **2008**, *64*, 339; (p) Yadav, J. S.; Reddy, B. V. S.; Geetha, V. Synth. Commun. **2003**, *33*, 717; (q) Rafiee, E.; Tangestaninejad, S.; Habibi, M. H.; Mirkhani, V. Bioorg. Med. Chem. Lett. 2004, 14, 3611; (r) Nagaraj, P.; Ramesh, N. G. Tetrahedron Lett. 2009, 50, 3970; (s) Watanabe, Y.; Itoh, T.; Sakakibara, T. Carbohydr. Res. 2009, 344, 516; (t) Gorityala, B. K.; Cai, S.; Ma, J.; Pasunooti, K. K.; Liu, X. W. Bioorg. Med. Chem. Lett. 2009, 19, 3093.
- (a) Kahne, D.; Walker, S.; Cheng, Y.; Engen, D. V. J. Am. Chem. Soc. 1989, 111, 6881; (b) Bolitt, V.; Chaguir, B.; Sinou, D. Tetrahedron Lett. 1992, 33, 2481; (c)

Houston, T. A.; Chervin, S. M.; Koreeda, M. ITE Lett. Batt. New Technol. Med. 2002, 3, 23.

- (a) Yadav, J. S.; Reddy, B. V. S.; Rao, C. V.; Chand, P. K.; Prasad, A. R. Synlett 2001, 1638; (b) Yadav, J. S.; Reddy, B. V. S. Synthesis 2002, 511; (c) Kawabata, H.; Kubo, S.; Hayashi, M. Carbohydr. Res. 2001, 333, 153. and references cited therein.
- (a) De Clercq, E.; Aerschot, A. V.; Herdewijin, P.; Baba, M.; Pauwels, R.; Balzarani, J. Nucleosides Nucleotides 1989, 8, 659; (b) Norbeck, D. W. Annu. Rep. Med. Chem. 1990, 25, 149; (c)Nucleosides and Nucleotides as Antitumor and Antiviral Agents; Chu, C. K., Baker, D. C., Eds.; Plenum: New York, 1993; (d) Dwek, R. A. Chem. Rev. 1996, 96, 683; (e) Varki, A. Glycobiology 1993, 3, 97; (f) Leutzinger, E. E.; Meguro, T.; Townsend, L. B.; Shuman, D. A.; Schweizer, M. P.; Stewart, C. M.; Robins, R. K. J. Org. Chem. 1972, 37, 3695; (g) Herscovici, J.; Montserret, R.; Antonakis, K. Carbohydr. Res. 1988, 176, 219; (h) Lee, K.; Choi, Y.; Gumina, G.; Zhou, W.; Schinazi, R. F.; Chu, C. K. J. Med. Chem. 2002, 45, 1313.
- Chandrasekhar, S.; Raji Reddy, Ch.; Chandrasekhar, G. Tetrahedron Lett. 2004, 45, 6481.
- 7. Colinas, P. A.; Bravo, R. D. Carbohydr. Res. 2007, 342, 2297.
- (a) Lorpitthaya, R.; Xie, Z. H.; Kuo, J. L.; Liu, X. W. Chem. Eur. J. 2008, 14, 1561;
  (b) Lorpitthaya, R.; Sophy, K. B.; Kuo, J. L.; Liu, X. W. Org. Biomol. Chem. 2009, 7, 1284;
  (c) Sudibya, H. G.; Ma, J.; Dong, X.; Ng, S.; Li, L-J.; Liu, X.-W.; Chen, P. Angew. Chem., Int. Ed. 2009, 48, 2723;
  (d) Lorpitthaya, R.; Xie, Z. H.; Sophy, K. B.; Kuo, J. L; Liu, X. W. Chem. Eur. J. 2010, 16, 588.
- Gorityala, B. K.; Lorpitthaya, R.; Bai, Y.; Liu, X. W. *Tetrahedron* 2009, 65, 5844.
  Sigma-Aldrich aluminium oxide, neutral, 150 mesh, STD grade was used for all reactions. Alumina was freshly prepared by heating at 200 °C under vacuum for 5 h and flushed with nitrogen. Alumina (5.3 equiv) was combined with 1 equiv of activated ZnCl<sub>2</sub> (heated under vacuum at 160 °C for 2 h), and to this mixture about 3 mL of dry THF or CH<sub>2</sub>Cl<sub>2</sub> was added with stirring for 5 min. The solvent was removed and the remaining solid was dried under vacuum.
- 11. Typical experimental procedure: To a stirred mixture of tri-O-acetyl-D-glucal (1) (100 mg, 0.37 mmol) and nitrogen nucleophile 2 (1 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (0.2 mL) was added 250 mg of ZnCl<sub>2</sub>/Al<sub>2</sub>O<sub>3</sub> at ambient temperature. The mixture was stirred for the appropriate amount of time (Table 2), and the extent of the reaction was monitored by TLC analysis. The reaction mixture was filtered and the solid residue was washed with CH<sub>2</sub>Cl<sub>2</sub> (5 mL). The combined filtrate was concentrated under vacuum to give the product which was purified by silica gel column chromatography (EtOAc/hexane = 1:2). The products were identified by IR, <sup>1</sup>H and <sup>13</sup>C NMR and mass spectroscopy. Spectroscopic data for selected products (major anomers): 4,6-di-O-acetyl-2,3-dideoxy-α-Derythro-hex-2-enopyranosyl-2'-nitrophenylsulfonamide (3c): <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): § 8.16-8.23 (m, 1H), 7.85-7.88 (m, 1H), 7.69-7.77 (m, 2H), 6.49 (d, J = 10.0 Hz, 1H), 5.76–5.96 (m, 2H), 5.67 (d, J = 9.2 Hz, 1H), 5.27 (d, J = 9.2 Hz, 1H), 4.10 (q, J = 6.8 Hz, 1H), 3.70-3.73 (m, 1H), 3.10 (d, J = 12.0 Hz, 1H), 2.07 (s, 3H), 2.03 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  170.4, 170.0, 135.2, 133.9, 133.2, 131.6, 131.1, 127.8, 126.0, 125.2, 125.1, 67.1, 64.1, 61.5, 20.9, 20.7; IR (CHCl<sub>3</sub>) 3285, 1738, 1539, 1367, 1236, 1170, 1032, 742 cm<sup>-1</sup>; HRMS (ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>16</sub>H<sub>19</sub>N<sub>2</sub>O<sub>9</sub>S: 415.0811, found: 415.0816. 4,6-Di-O-acetyl-2,3-dideoxy- $\alpha$ -D-erythro-hex-2-enopyranosyl-p-
- h) 5D1-0-detyle2,3-dutexys22-berylmo-lex22-enopyranosyt-pchlorophenylsulfonamide (3d): <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 7.88 (d, J = 6.9 Hz, 2H), 7.49 (d, J = 6.9 Hz, 2H), 5.94 (d, J = 6.9 Hz, 1H), 5.78 (d, J = 6.9 Hz, J = 7.8 Hz, 1H), 5.23 (dd, J = 8.7, 1.8 Hz, 1H), 3.93 (dd, J = 12.3, 3.6 Hz, 1H), 3.55-3.63 (m, 2H), 2.04 (s, 3H), 2.01 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz): δ 170.6, 170.1, 140.0, 139.4, 130.7, 129.3, 128.9, 128.7, 126.5, 67.2, 64.3, 61.9, 20.9, 20.7; IR (CHCl<sub>3</sub>): 3269, 1738, 1371, 1338, 1236, 1167, 1036, 756 cm<sup>-1</sup>; HRMS (ESI) m/z [M+H]\* calcd for C<sub>16</sub>H<sub>19</sub>CINO<sub>7</sub>S: 404.0571, found: 404.0574.
   4 6.Di-O-acetyl-2 3-dideoxy-a-p-enytran-ber-2-enopyranosyl-p-
- 4,6-Di-O-acetyl-2,3-dideoxy-α-ρ-erythro-hex-2-enopyranosyl-pfluorophenylsulfonamide (3e): <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 7.93–7.98 (m, 2H), 7.17–7.23 (m, 2H), 5.94 (d, J = 8.4 Hz, 1H), 5.79–5.84 (m, 2H), 5.62 (d, J = 5.7 Hz, 1H), 5.24 (dd, J = 9.0, 2.1 Hz, 1H), 3.94 (dd, J = 12.9, 4.5 Hz, 1H), 3.54–3.60 (m, 2H), 2.03 (s, 3H), 2.01 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz): δ 170.6, 170.0, 137.5, 130.6, 130.0, 129.9, 126.6, 116.4, 116.1, 67.1, 64.3, 61.9, 20.9, 20.7; IR (CHCl<sub>3</sub>): 3282, 1738, 1591, 1371, 1338, 1236, 1155, 1033, 754 cm<sup>-1</sup>; HRMS (ESI) *m/z* [M+H]\* calcd for C<sub>16</sub>H<sub>19</sub>FNO<sub>7</sub>S: 388.0866, found: 388.0869.
- 4, 6-Di-O-acetyl-2,3-dideoxy-α-D-erythro-hex-2-enopyranosyl-pmethoxyphenylsulfonamide (**3f**): <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 7.86 (d, *J* = 9.0 Hz, 2H), 6.97 (d, *J* = 9.0 Hz, 2H), 5.92 (d, *J* = 10.2 Hz, 1H), 5.68-5.82 (m, 2H), 5.59 (d, *J* = 7.5 Hz, 1H), 5.26 (dd, *J* = 8.7, 1.5 Hz, 1H), 3.90 (dd, *J* = 10.2, 3.0 Hz, 1H), 3.87 (s, 3H), 3.50-3.58 (m, 2H), 2.03 (s, 3H), 2.01 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz): δ 170.8, 170.2, 163.1, 133.1, 130.6, 129.6, 128.7, 126.8, 114.2, 66.8, 64.4, 62.0, 55.8, 21.0, 20.8; IR (CHCl<sub>3</sub>): 3420, 1647, 1238, 1165, 1034, 721 cm<sup>-1</sup>; HRMS (ESI) *m*/*z* [M+Na]<sup>+</sup> calcd for C<sub>17</sub>H<sub>21</sub>NO<sub>8</sub>SNa: 422.0886, found: 422.0888.
- 15. 4,6-Di-O-acetyl-2,3-dideoxy-α-*D*-erythro-hex-2-enopyranosyl-benzamide (**3i**): <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ 7.73 (d, *J* = 7.6 Hz, 2H), 7.45–7.48 (m, 1H), 7.36–7.40 (m, 2H), 6.51 (d, *J* = 10.0 Hz, 1H), 6.17 (d, *J* = 9.6 Hz, 1H), 5.83–5.93 (m, 2H), 5.26 (d, *J* = 9.6 Hz, 1H), 4.11–4.22 (m, 2H), 3.91–3.94 (m, 1H), 2.03 (s, 3H), 2.02 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): δ 170.9, 170.3, 166.7, 133.4, 132.2, 129.8, 129.6, 128.7, 127.2, 75.5, 74.3, 64.6, 63.1, 21.0, 20.9; IR (CHCl<sub>3</sub>): 3406, 1741, 1526, 1237, 1041 cm<sup>-1</sup>; HRMS (ESI) *m*/*z* [M+H]<sup>\*</sup> calcd for C<sub>17</sub>H<sub>20</sub>NO<sub>6</sub>: 334.1291, found: 334.1286.